

SHORT COMMUNICATION

BIOSYNTHESIS OF THE TROPINE MOIETY OF HYOSCYAMINE FROM δ -*N*-METHYLORNITHINE

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Abstract—The hyoscyamine and hyoscyne isolated from *Datura stramonium* plants which had been fed DL- α -*N*-methyl- ^{14}C -ornithine-2- ^{14}C were not radioactive. However, a significant incorporation of activity (0.63%) was found in hyoscyamine and hyoscyne obtained from plants which had been fed DL- δ -*N*-methyl- ^{14}C -ornithine-2- ^{14}C . A systematic degradation of the hyoscyamine indicated that all the activity was located in the tropine base at the bridgehead carbon C-1 (having the (R)-configuration) and on the *N*-methyl group. Furthermore, the ratio of activity at these two positions indicated that the δ -*N*-methylornithine had been incorporated without any cleavage of its *N*-methyl group.

NEUMAN and Schröter,¹ studying the metabolism of α - and δ -*N*-methyl- ^{14}C -ornithine in *Datura* spp., reported that the α -isomer yielded radioactive hyoscyamine and hyoscyne which were labelled on their *N*-methyl groups. However, the δ -*N*-methylornithine was a much poorer precursor of these alkaloids, and the activity was not confined to the *N*-methyl groups. From these results the authors claimed that α -*N*-methylornithine is a direct precursor of the pyrrolidine ring of tropine. It would thus follow that the nitrogen of the tropine nucleus is derived from the α -amino group of ornithine. However, Schütte and co-workers have recently shown^{2,3} that the tropine nitrogen is derived from the δ -amino group of ornithine by carrying out feeding experiments with ^{15}N -labelled ornithine. Similar conflicting results were reported on the biosynthesis of the pyrrolidine ring of nicotine.^{4,5} By the use of doubly labelled methylornithines we showed⁶ that δ -*N*-methylornithine and not α -*N*-methylornithine was a specific precursor of the pyrrolidine ring of nicotine. We considered that the methylornithines used by Neuman and Schroter were not authentic since they were prepared by an ambiguous method.⁷

We have now tested α - and δ -*N*-methyl- ^{14}C -ornithine-2- ^{14}C , prepared by unequivocal methods,⁶ as precursors of tropine. The labelled amino acids were added to the nutrient solution in which *Datura stramonium* plants were growing hydroponically. Neither isomer was absorbed rapidly from the nutrient solution; however, the δ -*N*-methylornithine was absorbed more quickly than the α -isomer. The alkaloids were isolated as previously described,⁸ inactive hyoscyamine and hyoscyne being added to facilitate separation and purification. The alkaloids obtained from plants which had been fed the α -*N*-methylornithine

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¹ D. NEUMAN and H.-B. SCHRÖTER, *Tetrahedron Letters* 1273 (1966).

² H. W. LIEBISCH and H. R. SCHÜTTE, *Z. Pflanzenphysiol.* **57**, 434 (1967).

³ H. W. LIEBISCH, A. S. RADWAN and H. R. SCHÜTTE, *Ann. Chem.* **721**, 163 (1969).

⁴ H.-B. SCHRÖTER and D. NEUMAN, *Tetrahedron Letters* 1279 (1966).

⁵ E. LEETE, E. G. GROS and T. J. GILBERTSON, *Tetrahedron Letters* **587** (1964).

⁶ T. J. GILBERTSON and E. LEETE, *J. Am. Chem. Soc.* **89**, 7085 (1967).

⁷ D. NEUMAN and H.-B. SCHRÖTER, *Z. Chem.* **5**, 385 (1965).

⁸ E. LEETE, *J. Am. Chem. Soc.* **84**, 55 (1962).

were inactive. The hyoscyamine and hyoscyne obtained from the plants fed δ -*N*-methylornithine were both radioactive and the incorporation of activity (0.63%) was significantly greater than the incorporation of ornithine-2- 14 C administered by the same methods to plants of a similar age (0.085%).⁹ The hyoscyamine was degraded systematically by the previously described stereospecific method which gave information on the activity at C-1 having the (R)-configuration.^{8,10} Activity at the *N*-methyl group of tropine was determined by the method of Brown and Byerrum.¹¹ Activities of the degradation products are recorded in Table 1. It was established that all the activity was located on the *N*-methyl group and at C-1. Furthermore the ratio of activity at C-1 to that of the *N*-methyl group of tropine was almost the same as the ratio of activity at C-2 to that of the *N*-methyl group in the administered δ -*N*-methyl- 14 C-ornithine-2- 14 C, indicating that the methylated amino acid was being incorporated intact without cleavage of the *N*-methyl group. Baralle and Gros,¹² comparing α - and δ -*N*-methyl- 3 H-ornithine, found that the δ -*N*-methylornithine was a much superior precursor of the *N*-methyl groups of cuscohygrine and hyoscyamine in *Atropa belladonna*.

In tobacco, δ -*N*-methylornithine, although serving as a precursor of the *N*-methylpyrrolidine of nicotine, is not a "normal" biosynthetic intermediate between ornithine and

TABLE 1. HYOSCYAMINE AND ITS DEGRADATION PRODUCTS

	Activity (dis/min/mM $\times 10^{-6}$)	Relative activity
Hyoscyamine hydrochloride	1.8	100
(\pm)-Tropidine methiodide	1.8	100
(+)- α -Methyltropidine dibenzoyl- <i>d</i> -tartrate	1.8	100
Cycloheptanone	1.55	86
1-Phenylcycloheptanol	1.53	85
Benzoic acid [C-1]	1.47	82
Triethylmethylammonium iodide [<i>N</i> -methyl]	0.15	8.3

Activity at C-1/Activity at *N*-methyl = 9.8.

nicotine. Thus ornithine-2- 14 C labels the pyrrolidine ring equally at C-2' and C-5', whereas δ -*N*-methylornithine-2- 14 C labels only C-2'. These results were rationalized⁶ by postulating that the tobacco plant is able to convert δ -*N*-methylornithine to *N*-methylputrescine with a non-specific decarboxylase.

In *Datura* the present results are consistent with δ -*N*-methylornithine being a real intermediate between ornithine and tropine, since it has previously been established that ornithine-2- 14 C labels tropine specifically at C-1. *N*-Methylputrescine¹³ and hygrine,^{14,15} have also been shown to be precursors of tropine. We thus favor the biosynthetic scheme illustrated in Fig. 1. 4-Methylaminobutanol- 14 C has been detected in *Datura* plants which were fed ornithine-2- 14 C.¹⁶

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¹⁰ E. LEETE, *Tetrahedron Letters* 1619 (1964).

¹¹ S. A. BROWN and R. U. BYERRUM, *J. Am. Chem. Soc.* **74**, 1523 (1952).

¹² F. E. BARALLE and E. G. GROS, *Chem. Commun.* 721 (1969).

¹³ H. W. LIEBISCH, W. MAIER and H. R. SCHÜTTE, *Tetrahedron Letters* 4079 (1966).

¹⁴ D. G. O'DONOVAN and M. F. KEOGH, *J. Chem. Soc. (c)* 223 (1969).

¹⁵ H. R. SCHÜTTE, *Abstracts Fifth IUPAC Symposium on the Chemistry of Natural Products*, p. 86, London (1968).

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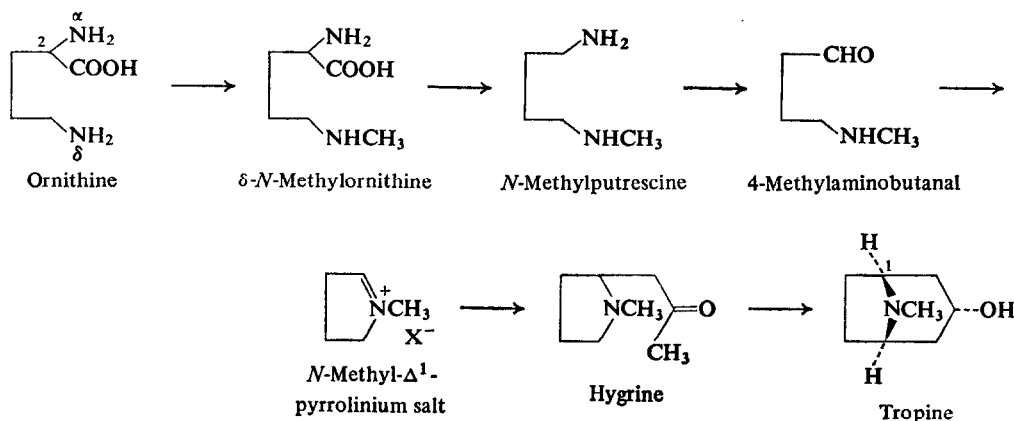


FIG. 1. BIOSYNTHETIC SCHEME FOR TROPINE.

EXPERIMENTAL

General Methods

Radioactivity measurements were carried out in a Nuclear Chicago liquid scintillation system, Model 724, using as solvents either toluene or dioxane, with the usual scintillators.¹⁷

 *α - And δ -*N*-methyl-¹⁴C-ornithine-2-¹⁴C*

These *N*-methylornithines were prepared as previously described.⁶ The actual weights and activities fed were:

α -N-Methylornithine. DL- α -*N*-methyl-¹⁴C-ornithine hydrochloride (135 mg), 4.1×10^6 dis/min. DL- α -*N*-methylornithine-2-¹⁴C hydrochloride (18 mg), 3.4×10^7 dis/min. Activity at C-2/Activity at *N*-methyl = 8.3.

δ -N-Methylornithine. DL- δ -*N*-methyl-¹⁴C-ornithine hydrochloride (14.5 mg), 3.0×10^7 dis/min. DL- δ -*N*-methylornithine-2-¹⁴C hydrochloride (138.5 mg), 3.15×10^8 dis/min. Activity at C-2/Activity at *N*-methyl = 10.5.

Feeding of the Tracers and Isolation of the Alkaloids

The *Datura* plants were 5 months old and growing in hydroponics at the time of feeding (December). The residual radioactivity in the nutrient solution to which the α -*N*-methyl-¹⁴C-ornithine-2-¹⁴C had been added was: 80% 1 week after feeding, 78% after 2 weeks, 38% after 3 weeks. Residual activity in the nutrient solution to which the δ -*N*-methyl-¹⁴C-ornithine-2-¹⁴C had been added was 78%, 47% and 1% after the same times. The plants were harvested 3 weeks after administering the tracers and worked up as previously described,⁸ inactive hyoscyamine hydrochloride (200 mg) and hyoscine (200 mg) being added as carriers. The alkaloids obtained from the plants which had been fed the α -*N*-methylornithine had negligible activity. From the plants which had been fed the δ -*N*-methylornithine, hyoscyamine hydrochloride (198 mg, 1.8×10^6 dis/min/mM) and hyoscine hydrochloride (188 mg, 1.8×10^6 dis/min/mM) were obtained. These activities indicate an absolute incorporation of radioactivity into the alkaloids of at least 0.63%.

The degradation of the hyoscyamine was carried out as previously described.⁸ Pyrolysis of the alkaloid yielded (\pm)-tropidine which was converted to its methiodide and subjected to a Hofmann degradation. The resultant (\pm)- α -methyltropidine was resolved with dibenzoyl-*D*-tartaric acid affording the (+)-isomer. Acid treatment followed by hydrogenation in the presence of Pd/C yielded cycloheptanone. Reaction with PhLi afforded 1-phenylcycloheptanol which was oxidized with KMnO₄ yielding benzoic acid. The hyoscyamine was demethylated with HI,¹² the resultant MeI being collected in an ethanolic solution of NEt₃ yielding MeNEt₃I. The activities of these degradation products are recorded in Table I.

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¹⁸ E. LEETE, *J. Am. Chem. Soc.* **82**, 612 (1960).